# 2003 Registration

The registration package includes morning and afternoon tea and lunch at the Convention Centre on Friday, morning and afternoon tea and a light lunch on Saturday. The conference dinner will take place at the University Staff Club on Saturday evening.

**Please send registration form with payment, and abstracts (if on disk), to the local organizer:**

**Dr David Saint**
**School of Molecular and Biomedical Sciences**
**University of Adelaide**
**SA 5005**
ph: 08 8303 3931 fax: 08 8303 3356 email: david.saint@adelaide.edu.au

**Closing date for Abstracts and Earlybird Registration is 31st July**

**Title:** **Name:**

**Institution:**

**Postal address:**

**phone:** **fax:**

**email:**

**Full Member of ASB** **$ 150**

**Student Member or Retiree** **$ 100**

**Non-Member** **$ 185**

Non-members may elect to use the extra registration charge towards payment of Society membership.

**Please tick box if you wish to join ASB** ☐

**Student Non-Member** **$ 124**

**Please tick box if you wish to join ASB** ☐

**Dinner on Saturday night (Sept 20, 2003) full payment** **$60**
**or student discount dinner** **$45**

**Surcharge for late registration (after 31 July)** **$15**

**Total Payment (included):**

**Make cheques payable to *Australian Society for Biophysics* OR**

**Please debit my Bankcard ☐ Mastercard ☐ Visa card ☐**

**Card No: _ _ _ _ _ _ _|_ _ _ ____|_ _ _ ____|_ _ _ ____** **Expiry Date:**

**Name on card:**

**Cardholder's Signature:**

**Abstract emailed / included on disk** ☐

**To be considered for presentation as: oral ☐ poster ☐ either ☐**

**I have applied for the Young Biophysicist Award** ☐

## Accommodation

There are a number of hotels and some budget accommodation within walking distance of the meeting.

The World Congress on Toxinology website has information on accommodation:

http://www.toxinology.net/ist_congress/men2sub4.htm

(you might find this doesn't work with some versions of Netscape)

Please make your own arrangements.

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## Abstract instructions:

Abstracts should fit in a rectangular box 175 mm wide by 120 mm high, with Times Roman font at 10 pt size. Title should be centred in all capitals, followed by a blank line, authors and their affiliations, blank line then text fully justified. Save as a word file (.doc) and email to David Saint, or save on disk and mail with registration.

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**MEASUREMENT OF LIPID-PROTEIN INTERACTIONS USING THE SURFACE PLASMON RESONANCE BIOSENSOR**

W.H. Sawyer and O.V. Rajaram
Russell Grimwade School of Biochemistry and Molecular Biology, University of Melbourne, Victoria 3010

Isolations from water and tubers collected from commercial potato washing plants showed that three *Erwinia* bacterium *Erwinia carotovora* subsp. *carotovora* (Ecc), *Erwinia carotovora* subsp. *atroseptica* (Eca) and *Erwinia chrysanthemi* (Echrys) caused tuber soft rot in South Australia. Fifty six South Australian isolates were sent to Scotland for identification, 77% of these *Erwinia* isolates were sourced from tubers, 18% from wash water samples and 5% from potato stems. Of the 43 *Erwinia* isolated from tubers 88.4% were identified as Ecc and 11.6% as Eca. Of the 10 isolates sourced from water samples 40% were identified as Ecc, 40% as Eca, 10% as E.chrys and 10% as other *Erwinia* species. All 3 isolates recovered from potato stems were identified as Eca. Four washing plants in South Australia were monitored to determine the level of *Erwinia* in wash water samples and to determine where tubers were becoming infected in the washing production line. Tubers were sampled from the main areas of the washing plant: the field bins, initial wash, tumbler, underneath final rinse sprays bars and from the end line. Collected tubers were induced to rot by maintaining them at 100% relative humidity for 3 days at 25?C.
After incubation tubers were assessed for incidence and severity of tuber soft rot on a scale from 0 to 5, where 0 = no signs of soft rot, 1 = lenticel infection only, 2 = less then ¼ surface area (SA) affected with soft rot, 3 = ¼ - ½ SA affected with soft rot, 4 = ½ - ¾ SA affected with soft rot and 5 = greater than ¾ SA affected by soft rot. Tubers were often contaminated in the field, with 73% of all field bin samples having a low level of rot, with the average incidence and severity of field bin tubers being 26% and 0.7 respectively. However most tubers became infected when they were immersed in water in the initial wash and tumbler region. Average initial wash soft rot incidences and severity were 72% and 2.3, respectively. Average incidence and severity of soft rot in tubers selected from the tumbler region were 87% and 2.8, respectively.